



<https://doi.org/10.59298/ROJESR/2025/4.3.1800>

Hormonal Response of the Reproductive System to Phytochemicals: Targeting Prolactin, Estrogen, and Gonadotropins for Fertility Regulation

Byaruhanga Abura G.

Faculty of Medicine Kampala International University Uganda

ABSTRACT

Phytochemicals, bioactive compounds derived from plants, are emerging as potent modulators of the endocrine system with implications for reproductive health. This review explores the mechanistic pathways through which specific phytochemicals influence the secretion and activity of key reproductive hormones such as prolactin, estrogen, luteinizing hormone (LH), and follicle-stimulating hormone (FSH), with a focus on their potential roles in fertility regulation. By examining evidence from in vitro studies, animal models, and clinical trials, we evaluate how phytoestrogens, lignans, flavonoids, and saponins interact with hormone receptors and signaling cascades. The review also discusses the dualistic effects of phytochemicals as both fertility enhancers and contraceptives, depending on dose, timing, and metabolic context. Furthermore, the safety, pharmacokinetics, and potential for integrating phytochemicals into fertility management protocols are critically addressed. This synthesis underscores the need for rigorous standardization and clinical validation to harness phytochemicals as complementary or alternative therapies in reproductive endocrinology.

Keywords: Phytochemicals, Fertility Regulation, Prolactin, Estrogen, Gonadotropins

INTRODUCTION

Fertility regulation is a central concern in reproductive medicine, encompassing both the enhancement of fertility for individuals facing subfertility or infertility, and the suppression of fertility for contraceptive purposes [1]. Traditional and complementary medicine systems across cultures have long employed plant-based remedies to influence reproductive functions [2]. In recent years, scientific interest in these traditional practices has surged, largely due to the growing body of evidence supporting the hormonal activity of plant-derived bioactive compounds, known collectively as phytochemicals [3]. These compounds include flavonoids, lignans, saponins, alkaloids, and phenolic acids, many of which are capable of interacting with human endocrine systems to modulate hormonal signaling pathways [4]. The reproductive endocrine system is intricately controlled by hormonal feedback loops involving the hypothalamus, pituitary gland, and gonads—collectively referred to as the hypothalamic-pituitary-gonadal (HPG) axis [5]. Within this system, the secretion and regulation of prolactin, estrogen, and gonadotropins (luteinizing hormone [LH] and follicle-stimulating hormone [FSH]) play pivotal roles in maintaining reproductive health [6]. Dysregulation of any of these hormones can result in menstrual irregularities, anovulation, infertility, or subfertility in women, and impaired spermatogenesis and reduced libido in men [7]. Phytochemicals exhibit both estrogenic and anti-estrogenic properties depending on their structural configuration and receptor affinity [8].

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Their ability to bind estrogen receptors, modulate dopamine pathways, or influence GnRH release enables them to influence the hormonal milieu of the reproductive system in diverse ways [9]. Some act as adaptogens, enhancing hormonal balance, while others exert specific hormonal suppression or stimulation, indicating potential use in both fertility treatments and contraceptive development [10]. The interest in phytochemicals for reproductive regulation is not solely pharmacological but also socio-cultural. In many low-resource settings, access to hormonal therapies is limited, and plant-based options offer a potentially accessible and culturally accepted alternative [11]. However, despite increasing scientific attention, the application of phytochemicals in clinical fertility management remains underdeveloped. This is largely due to inconsistencies in herbal product quality, limited standardization in dosing, and incomplete mechanistic understanding. This review aims to examine the hormonal response of the reproductive system to phytochemicals, with a particular focus on the modulation of prolactin, estrogen, and gonadotropins. Through analysis of in vitro experiments, in vivo studies, and clinical findings, we explore the mechanisms through which phytochemicals exert their effects, the dualistic potential for both fertility enhancement and suppression, and the current limitations that impede their integration into mainstream reproductive healthcare.

Phytochemicals and Prolactin Regulation

Prolactin is a peptide hormone secreted by lactotroph cells in the anterior pituitary gland [12]. While its primary function is to initiate and maintain lactation postpartum, prolactin also plays a crucial role in reproductive regulation [13]. Elevated levels of prolactin, known as hyperprolactinemia, can disrupt the pulsatile secretion of GnRH from the hypothalamus, leading to reduced LH and FSH secretion, menstrual irregularities, and anovulatory cycles in women [14]. In men, elevated prolactin levels are associated with hypogonadism and erectile dysfunction [15]. Phytochemicals capable of modulating prolactin levels offer a valuable avenue for correcting these hormonal imbalances. One of the most studied herbs in this context is *Vitex agnus-castus*, commonly known as chasteberry. Extracts from this plant contain flavonoids and iridoid glycosides, which have demonstrated dopaminergic activity [16]. By acting as agonists at dopamine D₂ receptors in the hypothalamus, these phytochemicals suppress prolactin release from the pituitary gland, thereby restoring normal gonadotropin secretion and ovulatory function [13]. Clinical studies have confirmed the efficacy of chasteberry in managing premenstrual syndrome (PMS), luteal phase defects, and mild hyperprolactinemia [17,18].

Other plants such as *Withania somnifera* (ashwagandha), *Lepidium meyenii* (maca), and *Panax ginseng* have also been evaluated for their effects on prolactin, with variable outcomes [19–21]. For instance, *Withania somnifera* has demonstrated normalization of elevated prolactin levels in animal models of stress-induced reproductive dysfunction [22]. The adaptogenic nature of these botanicals may contribute to endocrine equilibrium by regulating the hypothalamic-pituitary-adrenal (HPA) axis, which is closely interlinked with the HPG axis. It is important to note that the effect of phytochemicals on prolactin is not always suppressive [13]. Some compounds, particularly those found in fenugreek (*Trigonella foenum-graecum*), appear to stimulate prolactin secretion, and have been traditionally used to enhance lactation [23]. This illustrates the context-dependent nature of phytochemical action, which can vary based on compound concentration, individual hormonal status, and physiological conditions such as pregnancy or menopause.

Phytoestrogens and Estrogenic Modulation

Estrogens are critical regulators of the reproductive system, playing a central role in the menstrual cycle, ovulation, pregnancy, and maintenance of secondary sexual characteristics [24]. The estrogenic system is regulated via interactions with two main nuclear receptors—estrogen receptor alpha (ER α) and estrogen receptor beta (ER β)—and a membrane-bound G-protein-coupled estrogen receptor (GPER) [25]. Phytochemicals known as phytoestrogens can mimic or modulate the effects of endogenous estrogens through these receptors due to their structural similarity to estradiol [8].

Phytoestrogens encompass a variety of chemical classes, including isoflavones, lignans, coumestans, and flavonoids [26]. The most well-known phytoestrogens are isoflavones such as genistein and daidzein, predominantly found in soybeans (*Glycine max*), and lignans found in faxseed (*Linum usitatissimum*) [27]. These compounds exert either estrogenic or anti-estrogenic effects depending on circulating estrogen levels, the target tissue, and receptor subtype predominance. For example, while genistein binds both ER α and ER β , it shows a higher affinity for ER β , which is more abundantly expressed in the ovary, prostate, and vascular endothelium [28].

In conditions of estrogen deficiency such as menopause, phytoestrogens can exert weak estrogenic effects and potentially ameliorate symptoms such as hot flashes and bone loss [29]. On the contrary, in premenopausal women with normal or elevated estrogen levels, these compounds may compete with endogenous estrogen for receptor binding and exert anti-estrogenic effects, thereby influencing ovulation, menstrual cycle regularity, and overall

fertility [30]. Animal studies have provided strong evidence for the modulation of the estrous cycle by phytoestrogens. In rodents, genistein and daidzein administration has been shown to induce uterine proliferation, stimulate vaginal opening, and accelerate puberty onset, effects consistent with estrogen receptor activation [31–34]. However, high doses can result in prolonged estrous cycles, endometrial hyperplasia, and anovulation, highlighting the importance of dosage in determining physiological outcomes [35]. In human studies, dietary intake of phytoestrogens has shown variable effects on fertility. For instance, populations with high soy consumption such as in East Asia report both benefits (e.g., reduced menopausal symptoms) and concerns (e.g., altered menstrual cycle length) [36]. Interventional trials involving women undergoing in vitro fertilization (IVF) have demonstrated that soy isoflavones may enhance endometrial receptivity and implantation rates, while other studies suggest a potential reduction in FSH and LH levels, which may impair folliculogenesis in some cases [37]. The dualistic nature of phytoestrogens—serving as both agonists and antagonists of estrogen—depends significantly on endogenous hormonal milieu and receptor expression patterns [38]. This capacity for context-dependent modulation positions them as potential tools for individualized fertility regulation. However, long-term safety remains an area of concern, particularly regarding their impact on reproductive tract development during early life exposure and the risk of estrogen-sensitive cancers.

Gonadotropin Modulation by Plant Extracts

Gonadotropins, comprising luteinizing hormone (LH) and follicle-stimulating hormone (FSH), are glycoprotein hormones secreted by the anterior pituitary under the control of gonadotropin-releasing hormone (GnRH) from the hypothalamus [39, 40]. These hormones regulate gametogenesis, follicle maturation, ovulation, and steroid hormone production in both sexes [41]. Modulating their levels through phytochemicals represents a novel strategy for enhancing or inhibiting reproductive capacity. Several plant extracts have shown the capacity to influence gonadotropin secretion through direct pituitary stimulation or via modulation of the hypothalamic-pituitary axis. *Tribulus terrestris*, widely used in traditional Ayurvedic and Chinese medicine, is reputed for its aphrodisiac and fertility-enhancing effects [42]. Studies in male rats have demonstrated increased LH and testosterone levels following administration of *T. terrestris*, with concurrent improvements in sperm motility and count [43]. The active constituents, including protodioscin, may stimulate the hypothalamus to release GnRH, thus elevating pituitary gonadotropin output [42]. In females, *Asparagus racemosus* has demonstrated significant gonadotropin-modulating effects in experimental models [44]. Treatment with extracts from this plant led to increased FSH and LH levels, enhanced follicular development, and improved fertility indices [43]. These effects are attributed to steroidal saponins that modulate the release of GnRH and the responsiveness of the anterior pituitary to GnRH stimulation [6].

Mucuna pruriens, a leguminous plant rich in L-DOPA, has also gained attention for its reproductive effects. It acts via dopaminergic pathways that suppress prolactin and may indirectly support the restoration of normal LH and FSH secretion [45]. In male models, *M. pruriens* has improved testosterone production and sperm parameters, suggesting an overall enhancement of the hypothalamic-pituitary-gonadal axis [45]. Conversely, some phytochemicals have suppressive effects on gonadotropin secretion. Gossypol, a polyphenolic compound derived from cottonseed (*Gossypium* species), has been studied as a male contraceptive due to its inhibition of spermatogenesis and reduction in LH and FSH levels [46]. The suppression appears to occur through both direct testicular toxicity and central hypothalamic-pituitary interference. Other plants like Papaya and Neem extracts have similarly been shown to inhibit gonadotropin secretion and spermatogenesis [47]. The modulation of gonadotropin levels by phytochemicals underscores their dual potential as fertility enhancers or contraceptives. Importantly, the effects are highly dependent on the plant species, dosage, duration of exposure, and individual hormonal milieu. While fertility-boosting herbs may support patients undergoing assisted reproductive therapies or suffering from hypogonadism, those with anti-gonadotropic effects may offer plant-based contraceptive alternatives.

Dualistic Effects and Dose Dependency

One of the most compelling characteristics of phytochemicals in reproductive health is their dose-dependent and context-specific activity [48]. Unlike synthetic pharmaceuticals that generally act in a linear dose-response manner, many plant-derived compounds exhibit biphasic or even multiphasic effects [49]. This means that at lower doses, certain phytochemicals may promote fertility by enhancing hormonal activity, while at higher doses, they may exert anti-fertility effects or result in hormonal suppression. For instance, genistein, an isoflavone found in soy, has been widely studied for its effects on the reproductive system. At low concentrations, genistein can mimic endogenous estrogen, bind to estrogen receptors, and facilitate physiological responses such as endometrial thickening and follicular growth [50]. However, at higher doses or with prolonged exposure, genistein may exert anti-estrogenic

effects by competitively inhibiting the binding of natural estrogens to their receptors, potentially leading to menstrual disturbances or anovulation [51]. In some cases, high doses of genistein in neonatal rodents have resulted in reproductive tract abnormalities and long-term infertility, underlining the critical role of timing in exposure [52]. Similarly, saponins from *Tribulus terrestris* have been shown to stimulate LH secretion and enhance testosterone synthesis in men at moderate doses, thus improving spermatogenesis and libido [42]. However, excessive intake or prolonged use may disrupt the endocrine balance, leading to diminished returns or even suppression of hormonal function. In women, excessive estrogenic stimulation from phytoestrogens can lead to endometrial hyperplasia or irregular bleeding [53]. The dualistic action of phytochemicals can be attributed to their structural properties, receptor affinities, metabolism, and the physiological state of the individual. For example, in premenopausal women with adequate endogenous estrogen, phytoestrogens may exert antagonistic effects, whereas in postmenopausal women, the same compounds may have agonistic, hormone-balancing effects [54]. Additionally, individual differences in gut microbiota composition can alter the bioconversion and bioavailability of certain phytochemicals, resulting in variable hormonal responses [55]. This variation emphasizes the need for personalized approaches to phytochemical-based interventions in fertility regulation.

Clinical Applications and Challenges

Despite the promising findings from experimental studies, the clinical application of phytochemicals in fertility management remains limited and inconsistent. One of the primary reasons is the lack of standardization in herbal products. Unlike synthetic drugs, phytochemicals are derived from complex mixtures in plant extracts, where concentration and bioactivity can vary widely depending on species, cultivation conditions, harvesting time, and processing techniques [56]. Another major challenge is the lack of robust clinical trials that adhere to international standards of evidence-based medicine [57]. While traditional use and anecdotal evidence support the fertility-modulating potential of many herbs, few have undergone rigorous randomized controlled trials to evaluate their efficacy and safety in well-defined human populations. Studies that do exist often have small sample sizes, short durations, and methodological limitations that hinder generalizability.

Safety concerns also limit the widespread application of phytochemicals in reproductive health. Some compounds may exhibit endocrine-disrupting activity, reproductive toxicity, or hepatotoxicity at higher doses. For example, gossypol, although effective as a male contraceptive, was found to be associated with irreversible infertility and cardiac toxicity in some cases [58]. The long-term safety of commonly consumed phytoestrogens in relation to hormone-sensitive cancers, such as breast and endometrial cancers, is still under investigation. Regulatory frameworks for herbal products vary across countries, and most phytochemicals are marketed as dietary supplements rather than pharmaceuticals. This regulatory ambiguity leads to variability in quality, labeling, and claims, creating confusion among consumers and healthcare providers alike.

Future Perspectives

As interest in plant-based therapies for reproductive health continues to grow, future research must adopt a multidisciplinary approach combining molecular biology, pharmacology, clinical medicine, and traditional knowledge. Advanced analytical tools such as metabolomics, proteomics, and transcriptomics will be essential in elucidating the mechanisms of action of phytochemicals at a cellular and systemic level.

The development of organoid models, such as human endometrial or ovarian organoids, offers a promising platform for testing phytochemical effects in a physiologically relevant environment. These models can simulate menstrual cycle dynamics and hormonal responsiveness, providing insight into how phytochemicals influence reproductive tissues over time.

Another promising area lies in the integration of phytochemicals with assisted reproductive technologies (ART). Certain phytocompounds may enhance oocyte quality, sperm motility, or endometrial receptivity when used as adjuncts in in vitro fertilization protocols. However, their use in this context will require stringent validation to ensure that they do not interfere with hormonal protocols or embryonic development.

Personalized phytomedicine is also on the horizon. By analyzing individual hormonal profiles, genetic polymorphisms, and gut microbiome composition, clinicians may one day tailor phytochemical therapies for optimal fertility outcomes. Artificial intelligence and machine learning could assist in predicting individual responses and optimizing treatment regimens based on complex biological data.

CONCLUSION

Phytochemicals offer a promising frontier in fertility regulation through targeted modulation of prolactin, estrogen, and gonadotropins. However, the complexity of their effects warrants a cautious, evidence-based approach. Bridging ethnobotanical knowledge with modern endocrinology can lead to safe, effective fertility therapies rooted in nature.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

REFERENCES

1. Jain M, Singh M. Assisted Reproductive Technology (ART) techniques. StatPearls – NCBI Bookshelf. 2023. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK576409/>
2. Rahmawati N, Benjamin MAZ, Wahyuni TS, Sani MHM, Kadir AA, Abdullah MNH, et al. Medicinal plants from Southeast Asia affecting the reproductive system. Food Bioscience. 2024;63:105666. doi:10.1016/j.fbio.2024.105666
3. Alum EU. Role of phytochemicals in cardiovascular disease management: Insights into mechanisms, efficacy, and clinical application. Phytomedicine Plus, 2025; 5(1),100695. <https://doi.org/10.1016/j.phyplu.2024.100695>.
4. Egba, Simeon I., Okonkwo Chibuzo O, Omeoga Humphrey C and Ekong I E. Comparative modulation of the reproductive system by ethanol leaf extracts of *Asystasia gangetica* and *Anthocleista vogelii* in male Wistar rats. European Journal of Medical and Health Sciences. 2020; 2(3): 387-392
5. Berenbaum SA, Beltz AM, Corley R. The importance of puberty for adolescent development. Advances in Child Development and Behavior. 2015;53–92. doi:10.1016/bs.acdb.2014.11.002
6. Marques P, De Sousa Lages A, Skorupskaite K, Rozario KS, Anderson RA, George JT. Physiology of GnRH and gonadotrophin secretion. Endotext – NCBI Bookshelf. 2024. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK279070/>
7. Bendarska-Czerwińska A, Zmarzły N, Morawiec E, Panfil A, Bryś K, Czarniecka J, et al. Endocrine disorders and fertility and pregnancy: An update. Frontiers in Endocrinology. 2023;13. doi:10.3389/fendo.2022.970439
8. Ugwu, CE., Sure, SM., Dike, CC., Okpoga, NA and Egba, SI. Phytochemical and *in vitro* antioxidant activities of methanol leave extract of *Alternanthera basiliiana*. Journal of Pharmacy Research, 2018; 12(6): 835-839
9. Casteel CO, Singh G. Physiology, gonadotropin-releasing hormone. StatPearls – NCBI Bookshelf. 2023. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK558992/>
10. Chavda VP, Sonak SS, Balar PC, Vyas K, Palandurkar P, Mule K, et al. Reviving fertility: Phytochemicals as natural allies in the fight against non-genetic male infertility. Clinical Complementary Medicine and Pharmacology. 2024;4(1):100128. doi:10.1016/j.ccmp.2024.100128
11. Ndhlala AR, Amoo SO, Ncube B, Moyo M, Nair JJ, Van Staden J. Antibacterial, antifungal, and antiviral activities of African medicinal plants. In: Elsevier eBooks. 2013. p. 621–59. doi:10.1016/B978-0-12-405927-6.00016-3
12. Sadiq NM, Tadi P. Physiology, pituitary hormones. StatPearls – NCBI Bookshelf. 2023. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK557556/>
13. Egba, Simeon Ikechukwu., Okonkwo, Chibuzor Onyinye., Ogbodo, John Onyebuchi and Eme, Dike (2020) Contraceptive Potentials of *Alstonia boonei* via Stimulation of Increased Prolactin Synthesis in Experimental Female Wistar Rats. Trends Nat. Prod. Res, 2020; 1(1): 43-50
14. Crosignani PG. Management of hyperprolactinemic infertility. Middle East Fertility Society Journal. 2012;17(2):63–9. doi:10.1016/j.mefs.2012.04.003
15. Redman B, Kitchen C, Johnson KW, Bezwada P, Kelly DL. Levels of prolactin and testosterone and associated sexual dysfunction and breast abnormalities in men with schizophrenia treated with antipsychotic medications. Journal of Psychiatric Research. 2021;143:50–3. doi:10.1016/j.jpsychires.2021.08.022
16. Kamal N, Asni NSM, Rozlan INA, Azmi MAHM, Mazlan NW, Mediani A, et al. Traditional medicinal uses, phytochemistry, biological properties, and health applications of Vitex sp. Plants. 2022;11(15):1944. doi:10.3390/plants11151944
17. Puglia LT, Lowry J, Tamagno G. Vitex agnus-castus effects on hyperprolactinaemia. Frontiers in Endocrinology. 2023;14. doi:10.3389/fendo.2023.1269781
18. Kasim-Karakas SE, Mishra S. Botanical treatment for polycystic ovary syndrome. In: Elsevier eBooks. 2009. p. 317–32. doi:10.1016/B978-0-12-374228-5.00015-9
19. Chauhan S, Srivastava MK, Pathak AK. Effect of standardized root extract of ashwagandha (*Withania somnifera*) on well-being and sexual performance in adult males: A randomized controlled trial. Health Science Reports. 2022;5(4). doi:10.1002/hsr2.741

20. Del Carpio NU, Alvarado-Corella D, Quiñones-Laveriano DM, Araya-Sibaja A, Vega-Baudrit J, Monagas-Juan M, et al. Exploring the chemical and pharmacological variability of *Lepidium meyenii*: A comprehensive review of the effects of maca. *Frontiers in Pharmacology*. 2024;15. doi:10.3389/fphar.2024.1360422
21. Leung KW, St Wong A. Ginseng and male reproductive function. *Spermatogenesis*. 2013;3(3):e26391. doi:10.4161/spmg.26391
22. Azgomi RND, Zomorodi A, Nazemyieh H, Fazljou SMB, Bazargani HS, Nejatbakhsh F, et al. Effects of *Withania somnifera* on reproductive system: A systematic review of the available evidence. *BioMed Research International*. 2018;2018:1–17. doi:10.1155/2018/4076430
23. Reeder C, Legrand A, O'Connor-Von SK. The effect of fenugreek on milk production and prolactin levels in mothers of preterm infants. *Clinical Lactation*. 2013;4(4):159–65. doi:10.1891/2158-0782.4.4.159
24. Delgado BJ, Lopez-Ojeda W. Estrogen. *StatPearls – NCBI Bookshelf*. 2023. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK538260/>
25. Biason-Laubert A, Lang-Muritano M. Estrogens: Two nuclear receptors, multiple possibilities. *Molecular and Cellular Endocrinology*. 2022;554:111710. doi:10.1016/j.mce.2022.111710
26. Bopana N, Saxena S. *Asparagus racemosus*—Ethnopharmacological evaluation and conservation needs. *Journal of Ethnopharmacology*. 2007;110(1):1–15. doi:10.1016/j.jep.2007.01.001
27. Křížová L, Dadáková K, Kašparovská J, Kašparovský T. Isoflavones. *Molecules*. 2019;24(6):1076. doi:10.3390/molecules24061076
28. Paterni I, Granchi C, Katzenellenbogen JA, Minutolo F. Estrogen receptors alpha (ER α) and beta (ER β): Subtype-selective ligands and clinical potential. *Steroids*. 2014;90:13–29. doi:10.1016/j.steroids.2014.06.012
29. Sunita P, Pattanayak S. Phytoestrogens in postmenopausal indications: A theoretical perspective. *Pharmacognosy Reviews/Bioinformatics Trends/Pharmacognosy Review*. 2011;5(9):41. doi:10.4103/0973-7847.79098
30. SI Egba, I D Udom and C O Okonkwo. Comparative Effect of Oral Administration of Some Dietary Lipids on Fertility Hormones of Female Wistar Albino Rats. *Global Journal of Biotechnology and Biochemistry*, 2014; 9(1): 24–29
31. Dinsdale EC, Chen J, Ward WE. Early life exposure to isoflavones adversely affects reproductive health in first but not second generation female CD-1 mice. *Journal of Nutrition*. 2011;141(11):1996–2002. doi:10.3945/jn.111.142281
32. Egba, Simeon I., Sunday, Godwin I and Anaduaka, Emeka G. The effect of oral administration of aqueous extract of *Newbouldia laevis* leaves on fertility hormones of male albino rats. *IOSR Journal of Pharmacy and Biological Sciences*, 2014; 9(3): 61–64
33. Li R, Zowalaty AEE, Chen W, Dudley EA, Ye X. Segregated responses of mammary gland development and vaginal opening to prepubertal genistein exposure in Bsl2 $^{-/-}$ female mice with lipodystrophy. *Reproductive Toxicology*. 2014;54:76–83. doi:10.1016/j.reprotox.2014.10.023
34. Dinsdale EC, Ward WE. Early exposure to soy isoflavones and effects on reproductive health: A review of human and animal studies. *Nutrients*. 2010;2(11):1156–87. doi:10.3390/nu2111156
35. Canivenc-Lavier MC, Bennetau-Pelissero C. Phytoestrogens and health effects. *Nutrients*. 2023;15(2):317. doi:10.3390/nu15020317
36. Messina M, Duncan A, Messina V, Lynch H, Kiel J, Erdman JW. The health effects of soy: A reference guide for health professionals. *Frontiers in Nutrition*. 2022;9. doi:10.3389/fnut.2022.970364
37. Rizzo G, Feraco A, Storz MA, Lombardo M. The role of soy and soy isoflavones on women's fertility and related outcomes: An update. *Journal of Nutritional Science*. 2022;11. doi:10.1017/jns.2022.15
38. Mueller SO. Phytoestrogens and their human metabolites show distinct agonistic and antagonistic properties on estrogen receptor (ER α) and ER β in human cells. *Toxicological Sciences*. 2004;80(1):14–25. doi:10.1093/toxsci/kfh147
39. Nedresky D, Singh G. Physiology, luteinizing hormone. *StatPearls – NCBI Bookshelf*. 2022. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK539692/>
40. Egba S I., Omodamiro, Olorunsola D., Obike, J C and Ali, S E. Influence on some female fertility hormonal response in wistar albino rats: Possible contraceptive role for methanol leaf extract of *Ocimum gratissimum*? *Journal of Chemical and Pharmaceutical Research*, 2015; 7(5): 889–898

41. Holesh JE, Bass AN, Lord M. Physiology, ovulation. StatPearls – NCBI Bookshelf. 2023. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK441996/>
42. Saeed M, Munawar M, Bi JB, Ahmed S, Ahmad MZ, Kamboh AA, et al. Promising phytopharmacology, nutritional potential, health benefits, and traditional usage of *Tribulus terrestris* L. herb. *Heliyon*. 2024;10(4):e25549. doi:10.1016/j.heliyon.2024.e25549
43. Egba, SI., Omeoga, HC., Oriaku CE., Jacobs EC., Nnabugwu FC., Lazarus JC and Echem EN. Down-regulatory Influence of Methanol and Aqueous Root Extracts of *Sphenocentrum jollyanum* on Some Fertility Hormones and the Effect on Testicular Size of Wistar Albino Rats. *Annual Research and Review in Biology*, 2017; 18(3): 1-8
44. Erten F, Er B, Ozmen R, Tokmak M, Gokdere E, Orhan C, et al. Effects of integrated extracts of *Trigonella foenum-graecum* and *Asparagus racemosus* on hot flash-like symptoms in ovariectomized rats. *Antioxidants*. 2025;14(3):355. doi:10.3390/antiox14030355
45. Shukla KK, Mahdi AA, Ahmad MK, Shankhwar SN, Rajender S, Jaiswar SP. *Mucuna pruriens* improves male fertility by its action on the hypothalamus–pituitary–gonadal axis. *Fertility and Sterility*. 2008;92(6):1934–40. doi:10.1016/j.fertnstert.2008.09.045
46. Coutinho EM. Gossypol: a contraceptive for men. *Contraception*. 2002;65(4):259–63. doi:10.1016/S0010-7824(02)00294-9
47. Satriyasa BK, Mahendra AN, Arijana IGK, Ruspawan DM. Unripe papaya seed ethanol extract (*Carica papaya* Linn.) inhibits FSH and LH of male mice (*Mus musculus*). *Biomedical & Pharmacology Journal*. 2018;11(2):979–84. doi:10.13005/bpj/1457
48. Lambert MR, Edwards TM. Hormonally active phytochemicals and vertebrate evolution. *Evolutionary Applications*. 2017;10(5):419–32. doi:10.1111/eva.12469
49. Calabrese EJ. Biphasic dose responses in biology, toxicology and medicine: Accounting for their generalizability and quantitative features. *Environmental Pollution*. 2013;182:452–60. doi:10.1016/j.envpol.2013.07.046
50. Di X, Yu L, Moore AB, Castro L, Zheng X, Hermon T, et al. A low concentration of genistein induces estrogen receptor- α and insulin-like growth factor-I receptor interactions and proliferation in uterine leiomyoma cells. *Human Reproduction*. 2008;23(8):1873–83. doi:10.1093/humrep/den087
51. Zin SRM, Omar SZ, Khan NLA, Musameh NI, Das S, Kassim NM. Effects of the phytoestrogen genistein on the development of the reproductive system of Sprague Dawley rats. *Clinics*. 2013;68(2):253–62. doi:10.6061/clinics/2013(02)oa21
52. Meena R, Supriya Ch, Reddy KP, Reddy PS. Altered spermatogenesis, steroidogenesis and suppressed fertility in adult male rats exposed to genistein, a non-steroidal phytoestrogen during embryonic development. *Food and Chemical Toxicology*. 2016;99:70–7. doi:10.1016/j.fct.2016.11.020
53. Singh G, Cue L, Puckett Y. Endometrial hyperplasia. StatPearls – NCBI Bookshelf. 2024. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK560693/>
54. Rietjens IMCM, Louisse J, Beekmann K. The potential health effects of dietary phytoestrogens. *British Journal of Pharmacology*. 2016;174(11):1263–80. doi:10.1111/bph.13622
55. Boronat A, Rodriguez-Morató J, Serreli G, Fitó M, Tyndale RF, Deiana M, et al. Contribution of biotransformations carried out by the microbiota, drug-metabolizing enzymes, and transport proteins to the biological activities of phytochemicals found in the diet. *Advances in Nutrition*. 2021;12(6):2172–89. doi:10.1093/advances/nmab085
56. Alum EU, Tufail T, Uti DE, Aja PM, Ofor CE, Ibiam UA, Ukaidi CUA, Alum BN. Utilizing Indigenous Flora in East Africa for Breast Cancer Treatment: An Overview. *Anticancer Agents Med Chem*. 2024 Sep 18. doi: 10.2174/0118715206338557240909081833.
57. Uti DE, Atangwho IJ, Alum EU, Egba SI, Ugwu OP-C, Ikechukwu GC. Natural Antidiabetic Agents: Current Evidence and Development Pathways from Medicinal Plants to Clinical use. *Natural Product Communications*. 2025;20(3). doi:10.1177/1934578X251323393
58. Waites N, Wang N, Griffin N. Gossypol: reasons for its failure to be accepted as a safe, reversible male antifertility drug. *Andrology*. 1998;21(1):8–12. doi:10.1046/j.1365-2605.1998.00092.x

CITE AS: Byaruhanga Abura G. (2025). Hormonal Response of the Reproductive System to Phytochemicals: Targeting Prolactin, Estrogen, and Gonadotropins for Fertility Regulation. Research Output Journal of Engineering and Scientific Research 4(3): 1-8. <https://doi.org/10.59298/ROJESR/2025/4.3.1800>